



BAX® System Real-Time PCR Assays

STEC Suite



QUA 18/11-12/20
ALTERNATIVE ANALYTICAL METHODS
FOR AGRIBUSINESS
<http://nf-validation.afnor.org/en>

Screening Assay - Part KIT2021 for *stx* and *eae*

KIT CONTENTS

96 PCR tubes with tablets (2 bags of 6 x 8 strips)
96 flat optical caps (12 x 8 strips)
1 bottle of protease (400 µL)
2 bottles of lysis buffer (12 mL)

Panel 1 Assay - Part KIT2008 for *E. coli* O26, O111, O121

KIT CONTENTS

48 PCR tubes with tablets (1 bag of 6 x 8 strips)
96 flat optical caps (12 x 8 strips)
1 bottle of protease (400 µL)
2 bottles of lysis buffer (12 mL)

Panel 2 Assay - Part KIT2009 for *E. coli* O45, O103, O145

KIT CONTENTS

48 PCR tubes with tablets (1 bag of 6 x 8 strips)
96 flat optical caps (12 x 8 strips)
1 bottle of protease (400 µL)
2 bottles of lysis buffer (12 mL)

INTENDED USE

Food processors and associated laboratories can use the BAX® System STEC suite as a quick and reliable method for detecting pathogenic Shiga toxin-producing *E. coli* (STEC) in a variety of foods, hemp and cannabis flowers. Each real-time PCR assay in the STEC suite is designed to report clear yes/no results at concentrations as low as 10⁴ cfu/mL after enrichment. The screening assay detects the STEC virulence genes (*stx*₁, *stx*₂ and *eae*) with a simple, single-stage enrichment; two panel assays can then use the same lysate to determine if an STEC-screening positive is also positive for one of the top six STEC serogroups (*E. coli* O26, O45, O103, O111, O121, and O145). With a processing time of approximately 55 minutes in the BAX® System Q7 instrument, the method returns results comparable to culture methods, but with a significantly faster time to result.

BAX® Systems are designed for use by qualified lab personnel who follow standard microbiology laboratory practice, including the safe handling and disposal of potentially pathogenic materials. The laboratory must comply with good laboratory practice (see ISO 7218 standard). **Field of use:** Data obtained from the BAX® System should not be used for human diagnostic or human treatment purposes. Equipment is not approved by the United States Food and Drug Administration or any other U.S or non-U.S. regulatory agency for use in human diagnostics or treatment. The BAX® System should not be used as the sole basis for assessing the safety of products for release to consumers. The information generated is only to be used in conjunction with the user's regular quality assurance program. Not approved for clinical diagnosis. Use for research and development, quality assurance and quality control under supervision of technically qualified persons.

PRINCIPLE OF THE METHOD

See the BAX® System User Guide for an overview of how the BAX® System method uses automated, real-time Polymerase Chain Reaction (PCR) technology.

MATERIALS

BAX® System Real-Time PCR Assays – STEC suite
STEC Screening assay for *stx* and *eae* (Part KIT2021);
STEC Panel 1 assay for *E. coli* O26, O111, O121 (Part KIT2008); or
STEC Panel 2 assay for *E. coli* O45, O103, O145 (Part KIT2009)

BAX® System start-up package. (equipment and supplies for up to 192 tests)

- BAX® System Q7 cyler/detector and computer workstation
- Heating blocks with inserts* capable of maintaining 37±2°C and 95±3°C
- Cooling blocks with inserts*
- PCR tube holder
- Capping/decapping tools
- Adjustable mechanical pipettes (5-50 µL; 20-200 µL)
- Repeating pipette
- Multi-channel pipette (8 channels – 5-50 µL)
- Cluster tubes with caps and racks
- Pipette tips with barriers
- Powder-free nitrile gloves

*The Automated Thermal Block (Catalog No. MCH2023) may be used in place of heating and cooling blocks.

Stomacher with bags.

Incubator capable of maintaining directed enrichment temperatures between 39-42°C and enrichment temperatures within ±2°C.

Note: Health Canada, AOAC, and AFNOR Certification standards require an incubator capable of maintaining ±1°C.

Enrichment media (See BAX® System User Guide for details)

- Tryptic soy broth (TSB) (Formulation dependent on matrix tested. For specific details, please reference the STEC protocols in the BAX® System User Guide)
- BAX® System MP media – Catalog No. MED2003 (2.5 kg)
- Buffered Peptone Water (BPW)
- Double Strength BPW

Note: For an NF-Validation method, please note that for the preparations of master solutions, you must follow the instructions from the EN ISO 6887 standards.

STORAGE AND SHELF LIFE

- Reagents and PCR tubes with tablets should be kept refrigerated at 2–8°C. Do not freeze.
- Reagents should be used by the expiration date stamped on the individual labels.
- After protease has been added to the lysis buffer, shelf life of the solution is 2 weeks when stored at 2-8°C.
- If storing PCR tubes with tablets in an open kit for more than 3 weeks, seal the Mylar bag of PCR tubes into a larger bag with desiccant or store at 4°C in a desiccation unit.

PRECAUTIONS

The BAX® System method includes sample enrichment procedures that nourish the growth of potential pathogens to detectable levels. Because pathogens can cause human illness, appropriate safety precautions must be taken when handling samples, media, reagents, glassware and other supplies and equipment that could be contaminated with potentially pathogenic bacteria.

Reagents used with the BAX® System assays should pose no hazards when used as directed. Before using this product, please review the Safety Data Sheets (SDS) included with your BAX® System purchase and also available at www.hygiena.com. Refer to your site practices for safe handling of materials at extreme temperatures.

SOFTWARE REQUIREMENTS

Before using this assay for the first time, install the most current version of the BAX® System software, then run a calibration report to check that “Real Time STEC Screen *stx*, *eae*”, “Real Time STEC Screen *stx* only”, “Real Time STEC Panel 1 O26, O111, O121” and “Real Time STEC Panel 2 O45, O103, O145” appears in the list of calibration files. See “Troubleshooting Calibration” in the BAX® System User Guide for details. If the report list does not contain these targets you must recalibrate the Q7 instrument to load the required dyes. Be sure to allow enough time to complete the calibration (about 1.5 to 2 hours) before starting the assay. For instructions and tips on calibrating the instrument, see the BAX® System User Guide.

ENRICHMENT PROTOCOL

1. Prepare Enrichment Broth

Prepare enrichment broth according to the manufacturer's instructions. See the BAX® System User Guide for common enrichment media recipes.

2. Collect and Enrich Samples

Method Approved by AOAC

- *Raw ground beef (375 g):* Homogenize sample with 1.5 L pre-warmed (46°C) glucose-containing TSB with 2mg/L novobiocin. Incubate at 41°C for 12-24 hours.
- *Raw ground beef with soy (325 g):* Homogenize sample with 975 mL pre-warmed (35°C) glucose-containing TSB with 10 g/L casamino acids and 8 mg/L novobiocin. Incubate at 41°C* for 12-24 hours.
- *Raw beef trim (325 g):* Gently massage sample with 975 mL room-temperature glucose-containing TSB with 10 g/L casamino acids and 8 mg/L novobiocin. Incubate at 41°C* for 15-24 hours.
- *Raw beef trim (375 g):* **TSB** – Gently massage sample with 1.5 L pre-warmed (46°C) glucose-containing TSB. Incubate at 41°C* for 12-24 hours. **MP Media** – Gently massage sample with 1.5 L pre-warmed (46°C) BAX® System MP media. Incubate at 41°C* for 12-24 hours.
- *Raw ground beef (25 g):* **BPW Media** - Homogenize sample with 225 mL pre-warmed (37°C) BPW. Incubate at 37°C for 10-24 hours. **mTSB Media** - Homogenize sample with 225 mL pre-warmed (37°C) mTSB + casamino acids. Incubate at 42°C for 10-24 hours.
- *Flour (25 g):* Homogenize sample with 225 mL pre-warmed (42°C) mTSB with 2mg/L novobiocin. Incubate at 42°C for 24 hours.

*Note: Incubation temperature must be maintained between 39 °C and 42 °C for this assay.

- *Dried cannabis and hemp flower:* Combine 10 g flower with 90 mL pre-warmed (37-42°C) BPW. Hand massage for 1 minute and incubate at 42°C for 22-26 hours.

- *Sampling cloths such as MicroTally® (375 g Beef Trim):* Combine one sampling cloth with 200 mL pre-warmed (42°C) MP media or mTSB + caa (modified TSB with casamino acids) media. Stomacher for 1 minute and incubate at 42 ± 1°C for 8-24 hours.

Method Approved by AFNOR

Test portions weighing more than 25 g have not been tested in the context of NF VALIDATION.

For preparation of initial suspensions, follow instructions of EN ISO 6887 standard.

- *Raw beef meats (25 g):* **BPW Media** - Homogenize sample with 225 mL pre-warmed (41.5°C) BPW. Incubate at 41.5°C for 10-24 hours.

MP Media - Homogenize sample with 225 mL pre-warmed (41.5°C) MP media. Incubate at 41.5°C for 7-24 hours.

- *Raw dairy products (25 g):* **Double strength (DS) BPW Media** - Homogenize sample with 225 mL pre-warmed (41.5°C) DS BPW. Incubate at 41.5°C for 20-24 hours.
- *Vegetables (25 g):* **MP Media** - Homogenize sample with 225 mL pre-warmed (41.5°C) MP media. Incubate at 41.5°C for 8-24 hours.

TEST PROTOCOL – ALL ASSAYS

3. Prepare Equipment

- 3.1 Turn on the heating blocks to 37°C and 95°C*.
- 3.2 Make sure cooling blocks are chilled to 2-8°C*.
**If using the Automated Thermal Block, follow the instructions in the Thermal Block User Guide for running the Gram Negative program.*
- 3.3 Power on the Q7 instrument and launch the BAX® System application.
- 3.4 Create a rack file (see User Guide for details).

IMPORTANT NOTE FOR STEC SCREENING WITH “STX ONLY”:
An alternative target drop-down option is available for running the “stx only” program. See the BAX® System User Guide for details.

4. Perform Lysis

- 4.1 Break cluster tubes apart.
- 4.2 Label and arrange cluster tubes in rack according to the rack file.
- 4.3 Prepare lysis reagent by adding 150 µL protease to one 12 mL bottle of lysis buffer.
- 4.4 Transfer 200 µL prepared lysis reagent to each cluster tube.
- 4.5 Transfer 20 µL enriched sample to the corresponding cluster tube.
- 4.6 Heat at 37°C for 20 minutes.
- 4.7 Heat at 95°C for 10 minutes.
- 4.8 Cool at 2-8°C for at least 5 minutes.

5. Hydrate PCR Tablets

- 5.1 Initialize the instrument by selecting RUN FULL PROCESS from the OPERATION menu.
- 5.2 Place a PCR tube rack onto a chilled (2-8°C) PCR cooling block.
- 5.3 Arrange strips of PCR tubes according to your rack file.
- 5.4 Remove the caps from the first strip of tubes with the decapping tool.
- 5.5 Transfer 30 µL lysate (from step 4.8) into PCR tubes, then seal with flat optical caps.
- 5.6 Repeat with remaining strips of PCR tubes until all PCR tablets have been hydrated.

Note: PCR tablets must be hydrated and re-sealed immediately after removing the caps from the PCR tubes.

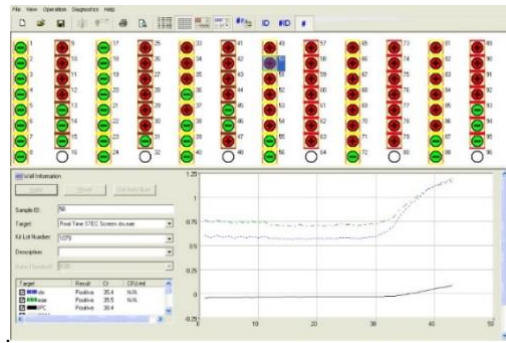
6. Amplify and Detect

- 6.1 At the “Ready for Rack Load” prompt, click the NEXT button and open the instrument drawer.
- 6.2 Place the rack of PCR tubes over the wells in the drawer, and check that the tubes are seated correctly.
- 6.3 Close the drawer and click the NEXT button to begin automated processing.

Note: If desired, remaining lysate can be sealed and stored at 2-8°C for additional testing with the BAX® System STEC suite and/or real-time *E. coli* O157:H7 assay.

7. Review Results

Qualitative results are displayed as a grid of color-cued icons in the top half of the screen:



	Green (-)	= Negative for target organism
	Red (+)	= Positive for target organism
	Yellow (?)	= Indeterminate result*
	Yellow (?) with red slash	= Signal error*

*Refer to the troubleshooting section in the User Guide for assistance.

Screening Assay Results (stx and eae)

Positive result – Indicates that both eae and stx are present in that sample. The amplification plot shows a rise in the stx (blue) and eae (green) targets.**

Negative result – Indicates that the combination of stx and eae is not present in that sample. If only one of the stx or eae targets is present, the sample is considered negative.

Screening Assay Results (“STX ONLY”)

Positive result – Indicates that stx is present in that sample. The amplification plot shows a rise in the stx (blue) target. The eae target is ignored.**

Negative result – Indicates that stx is not present in that sample. The eae target is ignored.

**Using the Test Protocol above, stored lysates of positive Screening samples can be run with the Panel 1 and Panel 2 assays to identify specific “Big 6” serogroup(s), if present.

Panel 1 Assay Results (*E. coli* O26, O111, O121)

Positive result – Indicates that one or more of the Panel 1 targets are present in that sample:

- *E. coli* O26 - the amplification plot shows a rise in the O26 (gold) target
- *E. coli* O111 - the amplification plot shows a rise in the O111 (grey) target
- *E. coli* O121 - the amplification plot shows a rise in the O121 (purple) target

Negative result – Indicates that none of the Panel 1 targets are present in that sample.

Panel 2 Assay Results (*E. coli* O45, O103, O145)

Positive result – Indicates that one or more of the Panel 2 targets are present in that sample:

- *E. coli* O45 - the amplification plot shows a rise in the O45 (magenta) target
- *E. coli* O103 - the amplification plot shows a rise in the O103 (brown) target
- *E. coli* O145 - the amplification plot shows a rise in the O145 (turquoise) target

Negative result – Indicates that none of the Panel 2 targets are present in that sample.

CONFIRMATION

Method Approved by AOAC

If desired, BAX@ method enriched samples can be confirmed with the reference culture method appropriate for the sample type, such as:

- U.S. FDA Bacteriological Analytical Manual (BAM)
- USDA FSIS Microbiology Laboratory Guidebook (MLG)
- Health Canada Compendium of Analytical Methods
- International Organization for Standardization (ISO)
- AOAC SMPR 2020.002

Method Approved by AFNOR

All samples identified as positive by the BAX@ System method must be confirmed in one of the following ways:

- Direct plating - Streak 10 µl of enrichment onto CT-SMAC and CHROMagar STEC. Incubate plates for 22-26 hours at 37°C. Test 1-5 typical colonies from at least one selective agar plate by:
 - Latex agglutination and BAX@ System Real Time STEC Screening assay or
 - BAX@ System Real Time STEC Screening assay and either BAX@ STEC Panel 1 or BAX@ STEC Panel 2 or BAX@ System Real-Time Assay for *E. coli* O157:H7.
- Immuno-concentration (IMS) - Perform IMS and streak 10 µl of enrichment onto CT-SMAC and CHROMagar STEC. Incubate plates for 22-26 hours at 37°C. Test 1-5 typical colonies from at least one selective agar plate by:
 - Latex agglutination and BAX@ System Real Time STEC Screening assay or
 - BAX@ System Real Time STEC Screening assay and either BAX@ STEC Panel 1 or BAX@ STEC Panel 2 or BAX@ System Real-Time Assay for *E. coli* O157:H7.

In the event of discordant results (presumptive positive by the alternative method and not confirmed by one of the means described above) the laboratory must follow the necessary steps to ensure the validity of the result obtained.

DISPOSAL

Decontaminate materials and dispose of biohazardous waste per your site practices and as required by federal, state and local regulations.

VALIDATION

The BAX@ System Real-Time PCR Assays for STEC (Screening, Panel 1 and Panel 2) have been certified by the AOAC Research Institute as Performance Tested MethodSM #091301. The performance of these assays was reviewed by AOAC-RI and was found to perform to the manufacturer’s specifications. Validation studies for foods and hemp and cannabis flowers, demonstrated BAX@ System sensitivity and specificity equal to or better than the reference culture based methods.

The USDA Food Safety and Inspection Service (USDA-FSIS) has adopted the BAX@ System STEC suite for monitoring meat products and carcass and environmental sponges. See FSIS Microbiology Laboratory Guidebook (MLG) Method #5B.04 for details and protocols. Please note that the enrichment and sample preparation protocols in the MLG may differ from those in the BAX@ System documentation.

The BAX@ System Real-Time PCR Assay Suite for STEC has been certified #QUA 18/11-12/20 according to NF VALIDATION rules. Validation studies conducted according to EN ISO 16140-2 standard found this test kit’s performance to satisfy the NF VALIDATION rules for raw beef meats, raw dairy products and vegetables.

The version of software used during the NF Validation study is indicated in the NF VALIDATION certificate of the alternative method. For more information, including validity dates, please refer to certificate QUA 18/11-12/20 available at <http://nf-validation.afnor.org>.

TECHNICAL ASSISTANCE

For questions or comments, please contact your local distributor. You can also call 800-863-6842 in the U.S., 1-302-695-5300 outside the U.S., or email diagnostics.support@hygiena.com.

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NOTICE: READ THIS LIMITATION OF WARRANTY AND LIABILITY BEFORE USING THE BAX@ SYSTEM EQUIPMENT, ASSAYS, AND/OR MEDIA (“BAX@ SYSTEM”). If the terms are not acceptable, notify Hygiena immediately and arrangements will be made for return of the unused Equipment, assays, and/or media to Hygiena and for the refund of the purchase price, less shipping costs. USE OF BAX@ SYSTEM EQUIPMENT, ASSAYS AND/OR MEDIA CONSTITUTES AN ACCEPTANCE OF ALL TERMS AND CONDITIONS OF THIS LIMITATION OF WARRANTY AND LIABILITY. Any additional or different terms in Buyer’s purchase form(s) are material alterations and hereby rejected.

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2. When used with BAX@ System assays, BAX@ System Equipment is warranted free of defects in materials, workmanship and design that may appear under normal and proper use within twelve (12) months from the installation date to the first end user. BAX@ System assays are warranted to conform to the assay description under the conditions of use specified in the user documentation to the expiration date stamped on the label. BAX@ System media is warranted to meet standard specifications in effect on the date of shipment. Hygiena MAKES NO OTHER WARRANTY, EITHER EXPRESS OR IMPLIED, INCLUDING, WITHOUT LIMITATION, ANY WARRANTY AGAINST INFRINGEMENT, ANY WARRANTY OF MERCHANTABILITY OR OF FITNESS FOR A PARTICULAR PURPOSE OR THOSE ARISING BY LAW, STATUTE, USAGE OF TRADE, OR COURSE OF DEALING. User assumes all risk and liability resulting from use of the BAX@ System Equipment, assays and media, whether used singly or in combination with other products.
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5. Externally caused failures, such as improper sample preparation, improper storage or loading of reagents, electrical outages, or out-of-specification environmental conditions are not covered under this warranty. Equipment failures caused by spills, abuse, misuse, negligence, or improper operation are not covered by this warranty. Modifications, service or repairs by parties other than Hygiena-authorized providers are not covered by this warranty and, in fact, void this warranty. Circumstances beyond the reasonable control of Hygiena, including fire, explosions, accidents, flood, labor trouble or shortage, war, act of or authorized by any government, inability to obtain suitable material, Equipment, fuel, power or transportation, or acts of God are not covered under this warranty.
6. The BAX@ System is designed to test only for the presence of the target organisms specified in the particular assay. The BAX@ System has been tested against many, but not all, strains of the target within the sample types specified in the user documentation. Hygiena, therefore, cannot and does not make any representation or warranty that the BAX@ System is capable of detecting every organism in the target genus, serotype, or species in any sample source. Accordingly, the BAX@ System should not be used as the sole test for the release of user’s products, nor should it be used as the sole basis for determining the safety of user’s products.
7. CUSTOMER/USER ASSUMES ALL RISKS IN USING THE BAX@ SYSTEM AND HYGIENA OR ITS AFFILIATES, DISTRIBUTORS, ITS LICENSORS OR REPRESENTATIVES SHALL HAVE NO LIABILITY TO CUSTOMER/USER OR TO ANY OTHER PERSON OR ENTITY FOR ANY INDIRECT, INCIDENTAL, SPECIAL, PUNITIVE, EXEMPLARY OR CONSEQUENTIAL DAMAGES WHATSOEVER, INCLUDING, BUT NOT LIMITED TO, LOSS OF REVENUE OR PROFIT, LOST OR DAMAGED DATA OR OTHER COMMERCIAL OR ECONOMIC LOSS EVEN IF CAUSED BY THE NEGLIGENCE OF HYGIENA OR ITS REPRESENTATIVES AND/OR IF HYGIENA HAS BEEN ADVISED OF THE POSSIBILITY OF SUCH DAMAGES, AND/OR IF THEY ARE FORESEEABLE.
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