



Prep Xpress: Best Practices for Tip Handling, Cleaning and Disposal

Introduction

The Prep Xpress User's Manual provides information on the proper disposable tips for use with the Prep Xpress instrument. In addition, the user's manual indicates the types of waste bags that can be used on the instrument. (The manual can be found on www.hygiena.com).

The following provides additional guidelines on Tip Handling, Cleaning and Disposal.

Waste Bin/Tip Disposal

Tips on Waste Bag Handling and Emptying

- Ensure the waste bag is fully seated and the sides of the bag are flush with the waste bin. When properly seated, a waste bag should be able to hold two (2) full racks of tips before needing to be disposed of and replaced with a new bag.
 - **Note:** Disposable tips are not heavy enough to fall to the bottom of the waste bag on their own. Manual intervention is necessary during waste bag installation to ensure tips fall to the bottom of the bag after instrument use.
- Empty the waste bin by removing the bin completely from the Prep Xpress instrument. Once the bin is outside the instrument, use gloved hands to touch only the outside of the bag and fold over the top of the bag to close it. Avoid touching the inside of the bag; remove and dispose of gloves when done.

Tip Pedestal/Rack Handling

Cleaning Options

Tip pedestals and racks may be:

- Removed and wiped down between use with 70 % alcohol (Reagent grade denatured alcohol, isopropanol or ethanol) or alternatively, non-bleach DNA removal solutions.
- Removed and dipped into non-bleach-containing cleaning solutions, followed by rinsing with water.
 - **Note:** Do not soak.
- Removed and placed in a UV light box for 30 min.

As a general rule of thumb:

- Prior to lysis (i.e., live bacteria), it is recommended to use 70% alcohol (70% Reagent grade denatured alcohol, isopropanol or ethanol) for cleaning.
- After lysis steps (i.e., dead bacteria/DNA), using non-bleach-containing alternatives for DNA removal is recommended.



Tip/Pellet Hydration

Hydration Pointers:

To ensure tips operate properly (avoid touching PCR pellets during hydration):

- Tap PCR tubes when loading each strip to ensure tablets are vertical in each tube.
- Confirm that the provided blue Eppendorf® blocks are being used on the instrument.

Tip Ejection Bar

Proper Plate Handling

Note that the ejection bar/plate handling area is a high-risk area for potential contamination. The recommendation is as follows:

- Avoid coming into contact with this area when removing plates. Pay special attention to lab coat sleeves, non-gloved exposed skin, etc.

Cleaning the Ejection Bar

Once samples are removed, the ejection bar can be cleaned in one of two ways:

- Wipe with 70% alcohol (70% ethanol, reagent-grade denatured alcohol or isopropanol)
- Wipe with a non-bleach-containing DNA removal solution (as an alternative to alcohol).

Routine Cleaning

General Cleaning

Recommendations are as follows:

- Disinfect/clean the tip pedestal/rack/dispensing area by wiping down all surfaces daily (end of the day).
- Clean any visible spills or residues as they occur to minimize contamination risk.

Technical Support

For assistance with Tip Handling and Disposal questions, please contact our technical support team at www.hygiena.com/support.